

Package ‘predmicror’

June 19, 2026

Title Fitting Predictive Microbiology Models

Version 1.3.2

Date 2026-06-05

Description Provides predictive microbiology model functions and convenience wrappers for fitting primary growth, microbial inactivation, dynamic, omnibus, and cardinal parameter models to experimental data using nonlinear least squares and related mixed-effects or time-varying workflows. Includes helper functions for extracting fitted values, calculating model diagnostics, and comparing fitted models. Implemented model families include those described by: Zwietering et al. (1990)

<doi:10.1128/AEM.56.6.1875-1881.1990>, Baranyi and Roberts (1994)

<doi:10.1016/0168-1605(94)90157-0>, Baranyi and Roberts (1995)

<doi:10.1016/0168-1605(94)00121-L>, Buchanan et al. (1997)

<doi:10.1006/fmic.1997.0125>, Richards (1959)

<doi:10.1093/jxb/10.2.290>, Fang et al. (2012)

<doi:10.1111/j.1750-3841.2012.02873.x>, Fang et al. (2013)

<doi:10.1016/j.fm.2012.12.005>, Huang (2008)

<doi:10.1111/j.1750-3841.2008.00785.x>, Huang (2009)

<doi:10.1016/j.jfoodeng.2008.07.011>, Huang (2013)

<doi:10.1016/j.foodcont.2012.11.019>, Geeraerd et al. (2005)

<doi:10.1016/j.ijfoodmicro.2004.11.038>, van Boekel (2002)

<doi:10.1016/S0168-1605(01)00742-5>, Peleg (1999)

<doi:10.1016/S0963-9969(99)00081-2>, Mafart et al. (2002)

<doi:10.1016/S0168-1605(01)00624-9>, Albert and Mafart (2005)

<doi:10.1016/j.ijfoodmicro.2004.10.016>, Rosso et al. (1993)

<doi:10.1006/jtbi.1993.1099>, Rosso et al. (1995)

<doi:10.1128/AEM.61.2.610-616.1995>, and Rosso et al. (1996)

<doi:10.4315/0362-028X-59.9.944>.

Depends R (>= 3.4.0)

Imports graphics, gslnls, nlme, Rdpack, stats, utils

RdMacros Rdpack

License GPL (>= 3)

LazyLoad yes

LazyData true

Encoding UTF-8

RoxygenNote 7.3.3

NeedsCompilation no

Suggests knitr, rmarkdown, htmltools, commonmark, shinyAce, testthat
(\geq 3.0.0), ggplot2, deSolve, FME, minpack.lm, reshape2,
readxl, shiny

VignetteBuilder knitr

URL <https://fsqanalytics.github.io/predmicror/>,
<https://github.com/fsqanalytics/predmicror>

BugReports <https://github.com/fsqanalytics/predmicror/issues>

Config/testthat/edition 3

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Repository CRAN

Date/Publication 2026-06-19 16:30:02 UTC

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aw	<i>Data of aw</i>
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Description

A dataset containing water activity and growth rate data

Usage

`data(aw)`

Format

A data frame with 9 rows and 3 variables:

aw Water activity

sqrtGR Square root of the growth rate

GR Growth rate

BaranyiFM

Baranyi and Roberts full growth model

Description

BaranyiFM function to fit the Baranyi & Roberts full growth model to a complete microbial growth curve. Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

BaranyiFM(t, Y0, Ymax, MUm_{ax}, lag)

Arguments

t	is a numeric vector indicating the time of the experiment
Y0	is the natural logarithm of the initial microbial concentration ($\ln(N_0)$) at time=0
Ymax	is the natural logarithm of the maximum concentration ($\ln(N_{\max})$) reached by the microorganism
MUm _{ax}	is the maximum specific growth rate given in time units
lag	is the duration of the lag phase in time units

Details

The model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the microbial concentration ($\ln(N(t))$) measured at time t.

Users should make sure that the microbial concentration input is entered in natural logarithm, $Y(t) = \ln(N(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Baranyi J, Roberts TA (1994). "A dynamic approach to predicting bacterial growth in food." *International Journal of Food Microbiology*, **23**, 277-294.

Examples

```
library(gslNls)
data(growthfull)
initial_values <- list(Y0 = -0.1, Ymax = 22, MUmmax = 1.7, lag = 5)
fit <- gsl_nls(lnN ~ BaranyiFM(Time, Y0, Ymax, MUmmax, lag),
  data = growthfull,
  start = initial_values
)
summary(fit)
```

BaranyiRM

Baranyi and Roberts reduced growth model

Description

BaranyiRM function to fit the Baranyi and Roberts growth model to a reduced microbial growth curve. Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
BaranyiRM(t, Y0, MUmmax, lag)
```

Arguments

t	is a numeric vector indicating the time of the experiment
Y0	is the natural logarithm of the initial microbial concentration ($\ln(N_0)$) at time=0
MUmmax	is the maximum specific growth rate given in time units
lag	is the duration of the lag phase in time units

Details

Model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the microbial concentration ($\ln(N(t))$) measured at time t.

Users should make sure that the microbial concentration input is entered in natural logarithm, $Y(t) = \ln(N(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez, <vcadavez@ipb.pt> and Ursula Gonzales-Barron, <ubarron@ipb.pt>

References

Baranyi J, Roberts TA (1995). "Mathematics of predictive microbiology." *International Journal of Food Microbiology*, **26**, 199-218.

Examples

```
## Example: Baranyi reduced model
library(gslnls)
data(growthred) # simulated data set.
initial_values <- list(Y0 = 0.1, MUmmax = 1.7, lag = 5) # define the initial values
fit <- gsl_nls(lnN ~ BaranyiRM(Time, Y0, MUmmax, lag),
  data = growthred,
  start = initial_values
)
summary(fit)
```

bias_factor

Bias and accuracy factors

Description

bias_factor() and accuracy_factor() calculate the multiplicative model bias and accuracy factors commonly used in predictive microbiology model validation.

Usage

```
bias_factor(observed, predicted)
```

```
accuracy_factor(observed, predicted)
```

Arguments

observed, predicted

Numeric vectors. Values must be positive.

Value

A numeric scalar.

Examples

```
observed <- c(7.0, 6.0, 5.0)
predicted <- c(7.1, 5.9, 5.2)
bias_factor(observed, predicted)
accuracy_factor(observed, predicted)
```

bixina	<i>Data concerning Staphylococcus aureus microbial inactivation in beef</i>
--------	---

Description

A dataset containing time (minutes), repetition and number of microorganisms (ln N).

Usage

```
data(bixina)
```

Format

A data frame with 18 rows and 3 variables:

Time Time in minutes

Rep Repetition

lnN Number of microorganism in ln scale

BuchananRM	<i>Buchanan reduced growth model</i>
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Description

BuchananRM function to fit the Buchanan reduced growth model to a reduced microbial growth curve. Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
BuchananRM(t, Y0, MUmax, lag)
```

Arguments

t	is a numeric vector indicating the time of the experiment
Y0	is the natural logarithm of the initial microbial concentration ($\ln(N_0)$) at time=0
MUm _{ax}	is the maximum specific growth rate given in time units
lag	is the duration of the lag phase in time units

Details

Model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the microbial concentration ($\ln(N(t))$) measured at time t.

Users should make sure that the microbial concentration input is entered in natural logarithm, $Y(t) = \ln(N(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez, <vcadavez@ipb.pt> and Ursula Gonzales-Barron, <ubarron@ipb.pt>

References

Buchanan RL, Whiting RC, Damert WC (1997). "When is simple good enough: a comparison of the Gompertz, Baranyi, and three-phase linear models for fitting bacterial growth curves." *Food Microbiology*, **14**(4), 313-326. ISSN 0740-0020, doi:10.1006/fmic.1997.0125.

Examples

```
## Example: Buchanan reduced model
library(gslnls)
data(growthred) # simulated data set.
initial_values <- list(Y0 = 0, MUmax = 1.7, lag = 5) # define the initial values
fit <- gsl_nls(lnN ~ BuchananRM(Time, Y0, MUmax, lag),
  data = growthred,
  start = initial_values
)
summary(fit)
```

CMAW

Cardinal model for water activity

Description

CMAW function to fit the water activity cardinal model (Rosso et al., 1993). Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
CMAW(x, AWmin, MUopt, AWopt)
```

Arguments

x	is a numeric vector indicating the water activity of the experiment
AWmin	is minimum water activity for growth
MUopt	is the optimum growth rate
AWopt	is optimum water activity for growth

Details

The model's inputs are:

x: Water activity

sqrtGR: the square root of the growth rate (h^{-1})

Users should make sure that the growth rate input is entered after a square root transformation, $\text{sqrGR} = \text{sqrt}(\text{GR})$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Rosso L, Lobry J, Charles (Bajard) S, Flandrois J (1995). "Convenient Model To Describe the Combined Effects of Temperature and pH on Microbial Growth." *Applied and environmental microbiology*, **61**, 610–6. doi:[10.1128/AEM.61.2.610616.1995](https://doi.org/10.1128/AEM.61.2.610-616.1995).

Examples

```
library(gslnls)
data(aw)
initial_values <- list(AWmin = 0.89, MUopt = 1.0, AWopt = 0.98)
fit <- gsl_nls(sqrtGR ~ CMAW(aw, AWmin, MUopt, AWopt),
  data = aw,
  start = initial_values
)
summary(fit)
```

CMInh

Cardinal model for growth inhibitors

Description

CMInh function to fit the growth inhibitors cardinal model (Rosso et al, 1993). Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
CMInh(x, MIC, MUopt, alpha)
```

Arguments

x	is a numeric vector indicating the inhibitor concentration of the experiment
MIC	is the minimum inhibitory concentration (mM or %, accordingly)
MUopt	is the optimum growth rate
alpha	is the shape parameter of the curve ($\alpha = 1$ the shape is linear; $\alpha > 1$ the shape is downward concave; and $\alpha < 1$ the shape is upward concave)

Details

The model's inputs are:

x: growth inhibitor concentration

sqrtGR: the square root of the growth rate ($time^{-1}$)

Users should make sure that the growth rate input is entered after a square root transformation, $\$sqrtGR = sqrt(GR)\$$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Rosso L, Lobry J, Charles (Bajard) S, Flandrois J (1995). "Convenient Model To Describe the Combined Effects of Temperature and pH on Microbial Growth." *Applied and environmental microbiology*, **61**, 610–6. doi:10.1128/AEM.61.2.610616.1995.

Examples

```
library(gslNls)
data(inh)
initial_values <- list(MIC = 0.89, MUopt = 1.0, alpha = 1)
fit <- gsl_nls(sqrtGR ~ CMInh(Conce, MIC, MUopt, alpha),
  data = inh,
  start = initial_values
)
summary(fit)
```

CMPH

Cardinal model for pH

Description

CMPH function to fit the pH cardinal model (Rosso et al, 1993). Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
CMPH(x, pHmax, pHmin, MUopt, pHopt)
```

Arguments

x	is a numeric vector indicating the pH of the experiment
pHmax	is the maximum pH for growth
pHmin	is the minimum pH for growth
MUopt	is the optimum growth rate
pHopt	is the optimum pH for growth

Details

The model's inputs are:

x: pH

sqrtGR: the square root of the growth rate ($time^{-1}$)

Users should make sure that the growth rate input is entered after a square root transformation, $sqrGR = \sqrt{GR}$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Rosso L, Lobry J, Charles (Bajard) S, Flandrois J (1995). “Convenient Model To Describe the Combined Effects of Temperature and pH on Microbial Growth.” *Applied and environmental microbiology*, **61**, 610–6. doi:10.1128/AEM.61.2.610616.1995.

Examples

```
library(gslNls)
data(ph)
initial_values <- list(pHmax = 9, pHmin = 3, MUopt = 1.0, pHopt = 7)
fit <- gsl_nls(sqrtGR ~ CMPH(pH, pHmax, pHmin, MUopt, pHopt),
  data = ph,
  start = initial_values
)
summary(fit)
```

CMTI

Cardinal model for temperature

Description

CMTI function to fit the temperature cardinal model (Rosso et al, 1993). Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
CMTI(x, Tmax, Tmin, MUopt, Topt)
```

Arguments

x	is a numeric vector indicating the temperature of the experiment
Tmax	maximum temperature for growth
Tmin	is minimum temperature for growth
MUopt	is the optimum growth rate
Topt	is optimum temperature for growth

Details

The model's inputs are:

x: Temperature

sqrtGR: the square root of the growth rate (h^{-1})

Users should make sure that the growth rate input is entered after a square root transformation, $\text{sqrtGR} = \text{sqrt}(\text{GR})$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Rosso L, Lobry J, Charles (Bajard) S, Flandrois J (1995). “Convenient Model To Describe the Combined Effects of Temperature and pH on Microbial Growth.” *Applied and environmental microbiology*, **61**, 610–6. doi:[10.1128/AEM.61.2.610616.1995](https://doi.org/10.1128/AEM.61.2.610616.1995).

Examples

```
library(gslnls)
data(salmonella)
initial_values <- list(Tmax = 42, Tmin = 1, MUopt = 1.0, Topt = 37)
fit <- gsl_nls(sqrtGR ~ CMTI(Temp, Tmax, Tmin, MUopt, Topt),
  data = salmonella,
  start = initial_values
)
summary(fit)

plot(salmonella$Temp, salmonella$sqrtGR^2)
lines(salmonella$Temp, fitted(fit)^2, col = "green")
plot(salmonella$Temp, salmonella$sqrtGR)
lines(salmonella$Temp, fitted(fit), col = "red")
```

compare_models

Compare fitted predmicror models

Description

compare_models() combines the output of `fit_metrics()` for two or more fitted models. It is useful when choosing between alternative primary growth, inactivation, or cardinal models fitted to the same response scale.

Usage

```
compare_models(..., sort_by = c("AIC", "BIC", "RMSE", "MAE", "none"))
```

Arguments

... predmicror_fit objects, or a single list of predmicror_fit objects.
 sort_by Character string. One of "AIC", "BIC", "RMSE", "MAE", or "none".

Value

A data frame with one row per fitted model.

Examples

```
data(growthfull)
huang <- fit_growth(
  growthfull,
  model = "HuangFM",
  time = "Time",
  response = "lnN",
  start = list(Y0 = 0, Ymax = 22, MUmmax = 1.7, lag = 5)
)
baranyi <- fit_growth(
  growthfull,
  model = "BaranyiFM",
  time = "Time",
  response = "lnN",
  start = list(Y0 = 0, Ymax = 22, MUmmax = 1.7, lag = 5)
)
compare_models(huang = huang, baranyi = baranyi)
```

dynamic_profile

Create a dynamic environmental profile

Description

dynamic_profile() stores time-varying environmental conditions such as temperature, pH, or water activity for dynamic predictive microbiology simulations. The profile is interpolated internally when dynamic models are solved between observed profile points.

Usage

```
dynamic_profile(time, temperature = NULL, ph = NULL, aw = NULL, ...)
```

Arguments

time	Numeric vector with profile times.
temperature	Optional numeric vector with temperatures at time.
ph	Optional numeric vector with pH values at time.
aw	Optional numeric vector with water activity values at time.
...	Additional named numeric vectors with the same length as time.

Value

A predmicror_dynamic_profile data frame sorted by time.

See Also

[predict_dynamic_growth\(\)](#), [predict_dynamic_inactivation\(\)](#)

Examples

```
profile <- dynamic_profile(
  time = c(0, 5, 10, 15),
  temperature = c(10, 4, 12, 20)
)
profile
```

dynamic_sensitivity *Finite-difference sensitivity for dynamic predictions*

Description

`dynamic_sensitivity()` perturbs parameters in a dynamic prediction and returns unscaled and scaled sensitivity coefficients. It is designed as a lightweight diagnostic for sampling design and parameter identifiability.

Usage

```
dynamic_sensitivity(
  type = c("growth", "inactivation"),
  profile,
  start,
  parameters = NULL,
  relative_delta = 1e-06,
  times = NULL,
  ...
)
```

Arguments

<code>type</code>	Character string. One of "growth" or "inactivation".
<code>profile</code>	A dynamic_profile() object or compatible data frame.
<code>start</code>	Named list of parameter values passed to the dynamic prediction function.
<code>parameters</code>	Character vector of parameter names to perturb. Defaults to all numeric scalar entries in <code>start</code> .
<code>relative_delta</code>	Relative perturbation size.
<code>times</code>	Optional output times.
<code>...</code>	Additional arguments passed to predict_dynamic_growth() or predict_dynamic_inactivation() .

Value

A data frame with time, parameter, prediction, sensitivity, and scaled sensitivity.

Examples

```

profile <- dynamic_profile(time = c(0, 5, 10), temperature = c(10, 15, 20))
sens <- dynamic_sensitivity(
  "growth",
  profile = profile,
  start = list(logN0 = 2, logNmax = 8, a = 0.08, Tmin = 7, lag = 1),
  times = seq(0, 10, by = 2),
  dt = 0.25
)
head(sens)

```

FangNLM

*Fang no lag growth model***Description**

FangNLM function to fit the Fang no lag growth model to an incomplete microbial growth curve. Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
FangNLM(t, Y0, Ymax, MUmax)
```

Arguments

t	is a numeric vector indicating the time of the experiment
Y0	is the natural logarithm of the initial microbial concentration ($\ln(N_0)$) at time=0
Ymax	is the natural logarithm of the maximum concentration ($\ln(N_{max})$) reached by the microorganism
MUmax	is the maximum specific growth rate given in time units

Details

Model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the microbial concentration ($\ln(N(t))$) measured at time t.

Users should make sure that the microbial concentration input is entered in natural logarithm, $Y(t) = \ln(N(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez, <vcadavez@ipb.pt> and Ursula Gonzales-Barron, <ubarron@ipb.pt>

References

Fang T, Gurtler JB, Huang L (2012). "Growth Kinetics and Model Comparison of Cronobacter sakazakii in Reconstituted Powdered Infant Formula." *Journal of Food Science*, **77**(9), E247-E255. doi:10.1111/j.17503841.2012.02873.x. Fang T, Liu Y, Huang L (2013). "Growth kinetics of Listeria monocytogenes and spoilage microorganisms in fresh-cut cantaloupe." *Food Microbiology*, **34**(1), 174-181. ISSN 0740-0020, doi:10.1016/j.fm.2012.12.005.

Examples

```
## Example: Fang no lag model
library(gslnls)
data(growthnolag) # simulated data set.
initial_values <- list(Y0 = 0, Ymax = 22, MUmmax = 1.7) # define the initial values
fit <- gsl_nls(lnN ~ FangNLM(Time, Y0, Ymax, MUmmax),
  data = growthnolag,
  start = initial_values
)
summary(fit)
```

 fit_cardinal

Fit a cardinal parameter model

Description

fit_cardinal() validates the input data, builds the nonlinear model formula, fits it with `gslnls::gsl_nls()`, and returns a `predmicror_fit` object.

Usage

```
fit_cardinal(data, model, x, response = "sqrtGR", start, ...)
```

Arguments

data	A data frame containing the environmental factor and response variables.
model	A model name, either quoted or unquoted. See <code>predmicror_models()</code> .
x	Column containing the environmental factor values, either quoted or unquoted.
response	Column containing the response values, either quoted or unquoted. Defaults to "sqrtGR".
start	Named list of initial parameter values for the selected model.
...	Additional arguments passed to <code>gslnls::gsl_nls()</code> .

Details

Cardinal models expect the response to be the square root of the growth rate, usually sqrtGR.

Value

A `predmicror_fit` object with the fitted model and metadata.

Examples

```
data(salmonella)
fit <- fit_cardinal(
  salmonella,
  model = "CMTI",
  x = "Temp",
  response = "sqrtGR",
  start = list(Tmax = 42, Tmin = 1, MUopt = 1, Topt = 37)
)
coef(fit)
```

fit_dynamic_growth *Fit dynamic microbial growth models*

Description

`fit_dynamic_growth()` estimates selected parameters of a dynamic Huang-type growth model by repeatedly solving the dynamic model and minimizing the residual sum of squares against observed data.

Usage

```
fit_dynamic_growth(
  data,
  profile,
  time,
  response,
  start,
  estimate = NULL,
  fixed = NULL,
  lower = NULL,
  upper = NULL,
  model = "huang",
  secondary = "huang_sqrt",
  scale = c("log10", "ln"),
  dt = 0.01,
  method = "rk4",
  temperature = "temperature",
  optimizer = "L-BFGS-B",
  control = list(),
  ...
)
```

Arguments

data	Data frame with observed microbial counts.
profile	A <code>dynamic_profile()</code> object or compatible data frame.
time, response	Column names in data containing observation time and microbial response.
start	Named list of starting parameter values passed to <code>predict_dynamic_growth()</code> . Numeric scalar entries are candidates for estimation.
estimate	Character vector of parameter names to estimate. If NULL, all numeric scalar entries in <code>start</code> except those listed in <code>fixed</code> are estimated.
fixed	Optional named list of parameters to keep fixed during fitting.
lower, upper	Optional named numeric vectors or lists with lower and upper bounds for estimated parameters.
model, secondary, scale, dt, method, temperature	Arguments passed to <code>predict_dynamic_growth()</code> .
optimizer	Optimization method passed to <code>stats::optim()</code> .
control	Optional control list passed to <code>stats::optim()</code> .
...	Additional arguments passed to <code>predict_dynamic_growth()</code> .

Value

A `predmicror_dynamic_fit` object.

Examples

```
profile <- dynamic_profile(time = c(0, 10), temperature = c(20, 20))
obs <- data.frame(time = c(0, 5, 10), logN = c(2, 3.3, 5.1))
fit <- fit_dynamic_growth(
  obs,
  profile = profile,
  time = "time",
  response = "logN",
  start = list(logN0 = 2, logNmax = 8, MUmax = 0.4, lag = 0),
  estimate = "MUmax",
  secondary = "constant",
  dt = 0.25
)
coef(fit)
```

fit_dynamic_inactivation

Fit dynamic microbial inactivation models

Description

`fit_dynamic_inactivation()` estimates selected parameters of a dynamic Weibull-Peleg inactivation model by minimizing the residual sum of squares against observed data.

Usage

```

fit_dynamic_inactivation(
  data,
  profile,
  time,
  response,
  start,
  estimate = NULL,
  fixed = NULL,
  lower = NULL,
  upper = NULL,
  model = "weibull_peleg",
  secondary = "constant",
  dt = 0.01,
  method = "rk4",
  temperature = "temperature",
  optimizer = "L-BFGS-B",
  control = list(),
  ...
)

```

Arguments

data	Data frame with observed microbial counts.
profile	A dynamic_profile() object or compatible data frame.
time, response	Column names in data containing observation time and microbial response.
start	Named list of starting parameter values passed to predict_dynamic_growth() . Numeric scalar entries are candidates for estimation.
estimate	Character vector of parameter names to estimate. If NULL, all numeric scalar entries in start except those listed in fixed are estimated.
fixed	Optional named list of parameters to keep fixed during fitting.
lower, upper	Optional named numeric vectors or lists with lower and upper bounds for estimated parameters.
model, secondary, dt, method, temperature	Arguments passed to predict_dynamic_inactivation() .
optimizer	Optimization method passed to stats::optim() .
control	Optional control list passed to stats::optim() .
...	Additional arguments passed to predict_dynamic_growth() .

Value

A `predmicror_dynamic_fit` object.

Examples

```

profile <- dynamic_profile(time = c(0, 10), temperature = c(60, 60))
obs <- data.frame(time = c(0, 5, 10), logN = c(7, 6, 5))
fit <- fit_dynamic_inactivation(
  obs,
  profile = profile,
  time = "time",
  response = "logN",
  start = list(logN0 = 7, b = 0.15, n = 1),
  estimate = "b",
  dt = 0.25
)
coef(fit)

```

fit_growth

Fit a primary growth model

Description

fit_growth() validates the input data, builds the nonlinear model formula, fits it with `gslnls::gsl_nls()`, and returns a `predmicror_fit` object.

Usage

```
fit_growth(data, model, time, response = "lnN", start, ...)
```

Arguments

data	A data frame containing the time and response variables.
model	A model name, either quoted or unquoted. See <code>predmicror_models()</code> .
time	Column containing time values, either quoted or unquoted.
response	Column containing the response values, either quoted or unquoted. Defaults to "lnN".
start	Named list of initial parameter values for the selected model.
...	Additional arguments passed to <code>gslnls::gsl_nls()</code> .

Details

Growth models expect the response to be the natural logarithm of the microbial concentration, usually $\ln N$.

Value

A `predmicror_fit` object with the fitted model and metadata.

Examples

```

data(growthfull)
fit <- fit_growth(
  growthfull,
  model = "HuangFM",
  time = "Time",
  response = "lnN",
  start = list(Y0 = 0, Ymax = 22, MUmmax = 1.7, lag = 5)
)
coef(fit)

```

fit_inactivation	<i>Fit a microbial inactivation model</i>
------------------	---

Description

fit_inactivation() validates the input data, builds the nonlinear model formula, fits it with [gslnls::gsl_nls\(\)](#), and returns a predmicror_fit object.

Usage

```
fit_inactivation(data, model, time, response = "logN", start, ...)
```

Arguments

data	A data frame containing the time and response variables.
model	A model name, either quoted or unquoted. See predmicror_models() .
time	Column containing time values, either quoted or unquoted.
response	Column containing the response values, either quoted or unquoted. Defaults to "logN".
start	Named list of initial parameter values for the selected model.
...	Additional arguments passed to gslnls::gsl_nls() .

Details

Inactivation models expect the response to be the base 10 logarithm of the microbial concentration, usually logN.

Value

A predmicror_fit object with the fitted model and metadata.

Examples

```

data(mafart2005Li11)
fit <- fit_inactivation(
  mafart2005Li11,
  model = "WeibullM",
  time = "Time",
  response = "logN",
  start = list(Y0 = 10, sigma = 3, alpha = 1)
)
coef(fit)

```

fit_metrics

Calculate model diagnostics for a fitted predmicror model

Description

fit_metrics() summarizes goodness-of-fit and information criteria for a predmicror_fit object. The metrics are calculated on the response scale used in the fitted model.

Usage

```

fit_metrics(object, ...)

## Default S3 method:
fit_metrics(object, ...)

## S3 method for class 'predmicror_fit'
fit_metrics(object, ...)

```

Arguments

object A predmicror_fit object.
... Currently unused. Included for future extension and S3 compatibility.

Value

A one-row data frame with columns: fit, model, type, response, response_scale, n (observations), p (parameters), SSE, RMSE, MAE, bias, RSE, R2, adj_R2, logLik, AIC, BIC, and converged.

Examples

```

data(growthfull)
fit <- fit_growth(
  data = growthfull,
  model = "HuangFM",
  time = "Time",
  response = "lnN",
  start = list(Y0 = 0, Ymax = 22, MUMax = 1.7, lag = 5)
)

```

```
)  
fit_metrics(fit)
```

fit_omnibus

Fit omnibus predictive microbiology models

Description

fit_omnibus() fits a nonlinear mixed-effects model in which the primary model is one of the parameterised predmicror primary models and one or more primary-model parameters are described by secondary covariate formulas.

Usage

```
fit_omnibus(  
  data,  
  type = c("growth", "inactivation"),  
  primary,  
  time,  
  response,  
  group,  
  secondary = NULL,  
  random,  
  correlation = NULL,  
  start,  
  method = "ML",  
  control = NULL,  
  ...  
)
```

```
fit_omnibus_growth(  
  data,  
  primary,  
  time,  
  response,  
  group,  
  secondary = NULL,  
  random,  
  correlation = NULL,  
  start,  
  method = "ML",  
  control = NULL,  
  ...  
)
```

```
fit_omnibus_inactivation(  
  data,
```

```

    primary,
    time,
    response,
    group,
    secondary = NULL,
    random,
    correlation = NULL,
    start,
    method = "ML",
    control = NULL,
    ...
  )

```

Arguments

data	A data frame.
type	Character string. One of "growth" or "inactivation".
primary	Character string naming a primary model registered in <code>predmicror</code> , for example "HuangNLM" or "WeibullM".
time, response, group	Column names for time, response, and grouping variable.
secondary	Optional named list of formulas, one per primary-model parameter. Parameters not listed are modelled as intercept-only effects.
random	Random-effects formula passed to <code>nlme::nlme()</code> . If the formula does not include a grouping term, <code> group</code> is added automatically.
correlation	Optional correlation structure. Use "AR1" for <code>nlme::corAR1()</code> within groups, NULL for none, or pass an <code>nlme</code> correlation object.
start	Numeric vector of starting values for fixed effects, in the order expected by <code>nlme::nlme()</code> .
method	Estimation method passed to <code>nlme::nlme()</code> .
control	Optional <code>nlme::nlmeControl()</code> object.
...	Additional arguments passed to <code>nlme::nlme()</code> .

Value

A `predmicror_omnibus_fit` object.

Examples

```

set.seed(1)
dat <- do.call(rbind, lapply(1:4, function(g) {
  Time <- c(1, 2, 4, 6, 8, 10)
  sigma <- 5 + 0.4 * g
  data.frame(
    Condition = g,
    Time = Time,
    Temp = 55 + g,

```

```

      logN = WeibullM(Time, Y0 = 7, sigma = sigma, alpha = 1.1) +
        rnorm(length(Time), 0, 0.03)
    )
  })
fit <- fit_omnibus_inactivation(
  dat,
  primary = "WeibullM",
  time = "Time",
  response = "logN",
  group = "Condition",
  secondary = list(sigma = ~ Temp),
  random = Y0 ~ 1,
  start = c(Y0 = 7, sigma = 1, sigma.Temp = 0.08, alpha = 1)
)
fit_metrics(fit)

```

 GeeraerdST

Geeraerd inactivation model

Description

GeeraerdST inactivation model for microbial inactivation curve. Returns the model parameters estimated according to data collected in microbial inactivation experiments.

Usage

```
GeeraerdST(x, Y0, Yres, kmax, S1)
```

Arguments

x	is a numeric vector indicating the heating time under a constant temperature of the experiment
Y0	is the initial (time=0) bacterial concentration ($\ln(N_0)$)
Yres	is a low asymptote reflecting the presence of a resistant sub-population ($\ln(N_{res})$)
kmax	is the maximum inactivation rate
S1	represents shoulder phase preceding the sharp inactivation slope of the curve

Details

The model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

N(t): the bacterial concentration measured at time t.

Users should make sure that the bacterial concentration input is entered in natural logarithm, $Y(t) = \ln(N(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Geeraerd AH, Valdramidis VP, Van Impe JF (2005). "GInaFiT, a freeware tool to assess non-log-linear microbial survivor curves." *International Journal of Food Microbiology*, **102**(1), 95-105. ISSN 0168-1605, doi:10.1016/j.ijfoodmicro.2004.11.038.

Examples

```
library(gslnls)
data(mafart2005Li11)
mafart2005Li11$lnN <- log(10) * mafart2005Li11$logN
initial_values <- list(Y0 = 18, Yres = 2, kmax = 0.7, S1 = 4)
fit <- gsl_nls(lnN ~ GeeraerdST(Time, Y0, Yres, kmax, S1),
  data = mafart2005Li11,
  start = initial_values
)
summary(fit)

plot(lnN ~ Time, data = mafart2005Li11)
lines(mafart2005Li11$Time, predict(fit), col = "blue")
```

growthfull

Data of a complete curve of microbial growth

Description

A dataset containing simulated data for a full growth model

Usage

```
data(growthfull)
```

Format

A data frame with 13 rows and 3 variables.

Time Time in minutes

logN Number of microorganism in log10 scale

lnN Number of microorganism in ln scale

growthnolag

Data of a no lag curve of microbial growth

Description

A dataset containing simulated data for a no lag growth model

Usage

```
data(growthnolag)
```

Format

A data frame with 10 rows and 3 variables.

Time Time in minutes

logN Number of microorganism in log10 scale

lnN Number of microorganism in ln scale

growthred

Data of a reduced curve of microbial growth

Description

A dataset containing simulated data for a reduced growth model

Usage

```
data(growthred)
```

Format

A data frame with 9 rows and 3 variables.

Time Time in minutes

logN Number of microorganism in log10 scale

lnN Number of microorganism in ln scale

HuangFM

Huang full growth model

Description

HuangFM function to fit the Huang full growth model to complete microbial growth curve. Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

HuangFM(t , Y_0 , Y_{max} , MU_{max} , lag)

Arguments

t	is a numeric vector indicating the time of the experiment
Y_0	is the natural logarithm of the initial microbial concentration ($\ln(N_0)$) at time=0
Y_{max}	is the natural logarithm of the maximum concentration ($\ln(N_{max})$) reached by the microorganism
MU_{max}	is the maximum specific growth rate given in time units
lag	is the duration of the lag phase in time units

Details

Model's inputs are:

t : time, assuming time zero as the beginning of the experiment.

$Y(t)$: the natural logarithm of the microbial concentration ($\ln(N(t))$) measured at time t .

Users should make sure that the microbial concentration input is entered in natural logarithm, $Y(t) = \ln(X(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez, <vcadavez@ipb.pt> and Ursula Gonzales-Barron, <ubarron@ipb.pt>

References

Huang L (2008). "Growth Kinetics of *Listeria monocytogenes* in Broth and Beef Frankfurters-Determination of Lag Phase Duration and Exponential Growth Rate under Isothermal Conditions." *Journal of Food Science*, **73**(5), E235-E242. doi:10.1111/j.17503841.2008.00785.x.

Examples

```
## Example: Huang full model
library(gslnls)
data(growthfull) # simulated data set.
initial_values <- list(Y0 = 0, Ymax = 22, MUmax = 1.7, lag = 5) # define the initial values
## Call the fitting function
fit <- gsl_nls(lnN ~ HuangFM(Time, Y0, Ymax, MUmax, lag),
  data = growthfull,
  start = initial_values
)
summary(fit)

confint(fit)

preds <- data.frame(predict(fit, interval = "prediction", level = 0.95))
plot(lnN ~ Time, data = growthfull, ylim = c(-1, 22))
lines(growthfull$Time, preds$fit, col = "blue")
lines(growthfull$Time, preds$upr, col = "red")
lines(growthfull$Time, preds$lwr, col = "red")
```

HuangNLM

*Huang no lag growth model***Description**

HuangNLM function to fit the Huang no lag growth model to an incomplete microbial growth curve. Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
HuangNLM(t, Y0, Ymax, MUmax)
```

Arguments

t	is a numeric vector indicating the time of the experiment
Y0	is the natural logarithm of the initial microbial concentration ($\ln(N_0)$) at time=0
Ymax	is the natural logarithm of the maximum concentration ($\ln(N_{max})$) reached by the microorganism
MUmax	is the maximum specific growth rate given in time units

Details

Model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the microbial concentration ($\ln(N(t))$) measured at time t.

Users should make sure that the microbial concentration input is entered in natural logarithm, $Y(t) = \ln(N(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez, <vcadavez@ipb.pt> and Ursula Gonzales-Barron, <ubarron@ipb.pt>

References

Huang (2013).

Examples

```
## Example: Huang no lag model
library(gslnls)
data(growthnolag) # simulated data set.
initial_values <- list(Y0 = 0, Ymax = 22, MUmmax = 1.7) # define the initial values
## Call the fitting function
fit <- gsl_nls(lnN ~ HuangNLM(Time, Y0, Ymax, MUmmax),
  data = growthnolag,
  start = initial_values
)
summary(fit)
```

HuangRGS

Huang reparameterized Gompertz survival model

Description

HuangRGS reparametrized Gompertz survival model for microbial inactivation. Returns the model parameters estimated according to data collected in microbial inactivation experiments.

Usage

HuangRGS(x, Y0, k, M)

Arguments

x	is a numeric vector indicating the heating time under a constant temperature of the experiment
Y0	is the initial microbial concentration (ln(cfu 1/g))
k	is the inactivation rate (1/s)
M	is a time constant (s)

Details

The model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the bacterial concentration ($\ln(X(t))$) measured at time t.

Users should make sure that the bacterial concentration input is entered in natural logarithm, $Y(t) = \ln(X(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Huang L (2009). "Thermal inactivation of *Listeria monocytogenes* in ground beef under isothermal and dynamic temperature conditions." *Journal of Food Engineering*, **90**(3), 380-387. ISSN 0260-8774, doi:[10.1016/j.jfoodeng.2008.07.011](https://doi.org/10.1016/j.jfoodeng.2008.07.011).

Examples

```
library(gslnls)
data(bixina)
initial_values <- list(Y0 = 5.6, k = 0.37, M = 6.8)
fit <- gsl_nls(lnN ~ HuangRGS(Time, Y0, k, M),
  data = bixina,
  start = initial_values
)
summary(fit)

plot(lnN ~ Time, data = bixina)
lines(bixina$Time, predict(fit), col = "blue")
```

HuangRM

Huang reduced growth model

Description

HuangRM function to fit the Huang reduced growth model to a reduced microbial growth curve. Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
HuangRM(t, Y0, MUmex, lag)
```

Arguments

t	is a numeric vector indicating the time of the experiment
Y0	is the natural logarithm of the initial microbial concentration ($\ln(N_0)$) at time=0
MUmax	is the maximum specific growth rate given in time units
lag	is the duration of the lag phase in time units

Details

Model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the microbial concentration ($\ln(N(t))$) measured at time t.

Users should make sure that the microbial concentration input is entered in natural logarithm, $Y(t) = \ln(N(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez, <vcadavez@ipb.pt> & Ursula Gonzales-Barron, <ubarron@ipb.pt>

References

Huang L (2008). "Growth Kinetics of *Listeria monocytogenes* in Broth and Beef Frankfurters-Determination of Lag Phase Duration and Exponential Growth Rate under Isothermal Conditions." *Journal of Food Science*, **73**(5), E235-E242. doi:10.1111/j.17503841.2008.00785.x.

Examples

```
## Example: Huang reduced model
library(gsl_nls)
data(growthred) # simulated data set.
initial_values <- list(Y0 = 0, MUmax = 1.7, lag = 5) # define the initial values
fit <- gsl_nls(lnN ~ HuangRM(Time, Y0, MUmax, lag),
  data = growthred,
  start = initial_values
)
summary(fit)
```

inh	<i>Data of INH antimicrobials</i>
-----	-----------------------------------

Description

A dataset containing antimicrobial concentration and growth rate data

Usage

```
data(inh)
```

Format

A data frame with 8 rows and 3 variables.

Conce Antimicrobial concentration

sqrtGR Square root of the growth rate

GR Growth rate

mafart2005Li11	<i>Data of microbial inactivation Albert and Mafart (2005)</i>
----------------	--

Description

A dataset containing time and log10 counts for microbial inactivation

Usage

```
data(mafart2005Li11)
```

Format

A data frame with 10 rows and 2 variables.

Time Time

logN Number of microorganism in log10 scale

omnibus_secondary	<i>Define an omnibus secondary model</i>
-------------------	--

Description

omnibus_secondary() defines a secondary model specification that can be used inside fit_omnibus(), fit_omnibus_growth(), or fit_omnibus_inactivation(). It allows a primary-model parameter to be described by another parameterised model already available in predmicror.

Usage

```
omnibus_secondary(model, x, transform = c("identity", "square"))
```

Arguments

model	Character string naming a registered predmicror model, for example "CMTI", "CMAW", "CMPH", or "CMInh".
x	Character string naming the covariate column used by the secondary model.
transform	Character string. One of "identity" or "square".

Value

An omnibus_secondary specification.

Examples

```
omnibus_secondary("CMTI", x = "Temp")
omnibus_secondary("CMTI", x = "Temp", transform = "square")
```

ph	<i>Data pH</i>
----	----------------

Description

A dataset containing pH and growth rate data

Usage

```
data(ph)
```

Format

A data frame with 14 rows and 3 variables.

pH pH of food

sqrtGR Square root of the growth rate

GR Growth rate

predict_dynamic_growth

Predict microbial growth under dynamic environmental conditions

Description

predict_dynamic_growth() solves a differential Huang-type growth model over a time-varying environmental profile. It is intended for forward prediction under dynamic temperature conditions and uses an internal fourth-order Runge-Kutta solver.

Usage

```
predict_dynamic_growth(
  profile,
  model = "huang",
  secondary = "huang_sqrt",
  start,
  times = NULL,
  scale = c("log10", "ln"),
  dt = 0.01,
  method = "rk4",
  temperature = "temperature"
)
```

Arguments

profile	A dynamic_profile() object or data frame with a time column and an environmental variable, usually temperature.
model	Character string. Currently only "huang" is supported.
secondary	Character string defining the secondary model for the growth rate. One of "huang_sqrt", "huang_full_sqrt", or "constant".
start	Named list of parameters. Use logN0 and logNmax for base-10 input, or Y0 and Ymax for natural-log input. For secondary = "huang_sqrt", supply a and Tmin. For secondary = "huang_full_sqrt", also supply b and Tmax. For secondary = "constant", supply MUmax.
times	Optional numeric vector of output times. If NULL, a regular sequence covering the profile is created from dt.
scale	Scale of the returned response column. One of "log10" or "ln".
dt	Maximum integration step used by the internal RK4 solver.
method	Solver method. Currently only "rk4" is implemented.
temperature	Name of the temperature column in profile.

Value

A `predict_dynamic_prediction` data frame with time, prediction, lnN, logN, temperature, and metadata attributes.

Examples

```

profile <- dynamic_profile(
  time = c(0, 5, 10, 15, 20),
  temperature = c(10, 4, 15, 15, 10)
)
pred <- predict_dynamic_growth(
  profile = profile,
  start = list(logN0 = 2, logNmax = 8.8, a = 0.0886, Tmin = 8.91, lag = 2),
  dt = 0.25
)
head(pred)

```

predict_dynamic_inactivation

Predict microbial inactivation under dynamic environmental conditions

Description

predict_dynamic_inactivation() solves a dynamic Weibull-Peleg type inactivation model over a time-varying environmental profile. The output is returned on the base-10 logarithmic scale.

Usage

```

predict_dynamic_inactivation(
  profile,
  model = "weibull_peleg",
  secondary = "constant",
  start,
  times = NULL,
  dt = 0.01,
  method = "rk4",
  temperature = "temperature"
)

```

Arguments

profile	A <code>dynamic_profile()</code> object or data frame with a time column.
model	Character string. Currently only "weibull_peleg" is supported.
secondary	Character string defining the secondary model for the Peleg rate parameter b. Use "constant" for constant conditions or "z_value" for a log-linear temperature effect.
start	Named list. Supply logN0, shape parameter n, and either b for secondary = "constant" or b_ref, T_ref, and z for secondary = "z_value".
times	Optional numeric vector of output times.
dt	Maximum integration step used by the internal RK4 solver.
method	Solver method. Currently only "rk4" is implemented.
temperature	Name of the temperature column in profile.

Value

A predmicror_dynamic_prediction data frame with time, logN, log_survival, temperature, and metadata attributes.

Examples

```
profile <- dynamic_profile(time = c(0, 10, 20), temperature = c(55, 58, 60))
pred <- predict_dynamic_inactivation(
  profile = profile,
  start = list(logN0 = 7, b_ref = 0.15, T_ref = 55, z = 8, n = 1.2),
  secondary = "z_value",
  dt = 0.25
)
head(pred)
```

predmicror_assistant *Assistant for predmicror*

Description

Answer questions about predmicror models using a deterministic local registry and, when available, an optional local Ollama model for prose. Model-fitting code is generated from package metadata and statically checked before being returned.

Usage

```
predmicror_assistant(
  query,
  model = "llama3-groq-tool-use:8b",
  root = NULL,
  data = NULL,
  file = NULL,
  sheet = NULL,
  sep = NULL,
  dec = ".",
  na.strings = c("", "NA"),
  task = NULL,
  time = NULL,
  response = NULL,
  temperature = NULL,
  ph = NULL,
  aw = NULL,
  inhibitor = NULL,
  return_context = FALSE,
  conversation = NULL,
  backend = c("auto", "ollama", "deterministic"),
  prefer_wrappers = TRUE,
```

```

    verify_code = TRUE,
    return_trace = FALSE
  )

```

Arguments

query	Character question to ask.
model	Ollama model name used when backend is "auto" or "ollama".
root	Path to the package root for context collection. Defaults to the installed package path when available, otherwise the current working directory.
data	Optional data frame to profile and use for data-aware code generation.
file	Optional path to a .csv, .txt, .tsv, .xls, or .xlsx file to read, profile, and use for data-aware code generation.
sheet	Optional Excel sheet name or index when file is .xls or .xlsx.
sep	Optional field separator for delimited text files. If NULL, a simple separator detector is used.
dec	Decimal mark for delimited text files.
na.strings	Character vector of strings to treat as missing values when reading delimited text files.
task	Optional data task override. Use one of "growth", "inactivation", or "cardinal"; NULL keeps automatic detection.
time, response, temperature, ph, aw, inhibitor	Optional column-name overrides used when data or file is supplied.
return_context	Logical; if TRUE, returns a list with answer and context.
conversation	Optional list or character vector with prior questions and answers to include as conversational context.
backend	Character string. One of "auto", "ollama", or "deterministic". "auto" uses Ollama when the CLI is available and otherwise falls back to deterministic registry-based output.
prefer_wrappers	Logical; if TRUE, generated fitting examples prefer <code>fit_growth()</code> , <code>fit_inactivation()</code> , and <code>fit_cardinal()</code> over direct <code>gsl_nls::gsl_nls()</code> calls.
verify_code	Logical; if TRUE, statically checks generated R code with <code>parse()</code> and simple model-scale/signature rules.
return_trace	Logical; if TRUE, returns prompt, backend, candidates, generated code, and validation metadata for debugging.

Value

Character response by default; list with answer and context when `return_context = TRUE` or list with trace when `return_trace = TRUE`.

Examples

```
predmicror_assistant("How do I fit a Huang model?", backend = "deterministic")
```

`predmicror_assistant_app`*Launch the predmicror assistant Shiny app*

Description

Starts the local assistant app bundled with the package. The app can read delimited text and Excel files, preview uploaded data, profile columns, apply optional manual column overrides, and pass the data profile to `predmicror_assistant()`. If the bundled app is not available, a fallback app is created from the installed R functions.

Usage

```
predmicror_assistant_app(  
  model = "llama3-groq-tool-use:8b",  
  root = NULL,  
  host = "127.0.0.1",  
  port = NULL,  
  launch.browser = interactive()  
)
```

Arguments

<code>model</code>	Default Ollama model name used by <code>predmicror_assistant()</code> . The app also exposes a model selector populated from <code>ollama list</code> when Ollama is available.
<code>root</code>	Optional package root used for assistant context collection.
<code>host</code>	Host passed to <code>shiny::runApp()</code> . Defaults to "127.0.0.1".
<code>port</code>	Optional port passed to <code>shiny::runApp()</code> .
<code>launch.browser</code>	Logical; whether to launch a browser.

Value

Runs the Shiny app.

Examples

```
predmicror_assistant_app()
```

predmicror_augment *Extract fitted values and residuals from a predmicror_fit*

Description

predmicror_augment() returns the original data, or optional new data, with columns containing fitted values and residuals. It is intentionally lightweight and does not require the broom package.

Usage

```
predmicror_augment(object, ...)

## Default S3 method:
predmicror_augment(object, ...)

## S3 method for class 'predmicror_fit'
predmicror_augment(object, newdata = NULL, ...)

## S3 method for class 'predmicror_fit'
as.data.frame(x, row.names = NULL, optional = FALSE, ...)
```

Arguments

object, x	A predmicror_fit object.
...	Additional arguments passed to predict().
newdata	Optional data frame used for prediction. If NULL, the data stored in the fitted object are used.
row.names, optional	Arguments required by the base as.data.frame() generic. They are accepted for method compatibility.

Value

A data frame containing the original columns plus .fitted, .resid when the response column is available, .model, and .type.

Examples

```
data(growthfull)
fit <- fit_growth(
  data = growthfull,
  model = "HuangFM",
  time = "Time",
  response = "lnN",
  start = list(Y0 = 0, Ymax = 22, MUmmax = 1.7, lag = 5)
)
head(predmicror_augment(fit))
```

```
predmicror_fit_methods
```

```
Methods for predmicror_fit objects
```

Description

These methods delegate to the fitted nonlinear least-squares object returned by `gslnls::gsl_nls()`, while preserving the model metadata added by the `fit_*`(`)` wrappers.

Usage

```
## S3 method for class 'predmicror_fit'
print(x, ...)

## S3 method for class 'predmicror_fit'
summary(object, ...)

## S3 method for class 'predmicror_fit'
predict(object, newdata = NULL, ...)

## S3 method for class 'predmicror_fit'
plot(
  x,
  xlab = x$x,
  ylab = paste0(x$response, " (", x$response_scale, ")"),
  ...
)

## S3 method for class 'predmicror_fit'
coef(object, ...)

## S3 method for class 'predmicror_fit'
fitted(object, ...)

## S3 method for class 'predmicror_fit'
residuals(object, ...)

## S3 method for class 'predmicror_fit'
vcov(object, ...)

## S3 method for class 'predmicror_fit'
logLik(object, ...)

## S3 method for class 'predmicror_fit'
AIC(object, ..., k = 2)

## S3 method for class 'predmicror_fit'
```

```
BIC(object, ...)
```

Arguments

x, object	A predmicror_fit object.
...	Additional arguments passed to the underlying method.
newdata	Optional data frame for prediction. If omitted, predictions are computed for the original data.
xlab, ylab	Axis labels used by plot().
k	Penalty used by AIC().

Value

The value returned by the corresponding method for the underlying nonlinear model object. plot() invisibly returns x.

predmicror_models *List models available through the fitting wrappers*

Description

List models available through the fitting wrappers

Usage

```
predmicror_models(type = "all")
```

Arguments

type	Character string. One of "all", "growth", "inactivation", or "cardinal".
------	--

Value

A data frame with the model type, model name, expected response scale, and parameters that must be supplied in start.

Examples

```
predmicror_models()
predmicror_models("growth")
```

predmicror_omnibus_methods

Methods for omnibus fits

Description

Methods for omnibus fits

Usage

```
## S3 method for class 'predmicror_omnibus_fit'  
print(x, ...)
```

```
## S3 method for class 'predmicror_omnibus_fit'  
summary(object, ...)
```

```
## S3 method for class 'predmicror_omnibus_fit'  
coef(object, ...)
```

```
## S3 method for class 'predmicror_omnibus_fit'  
fitted(object, ...)
```

```
## S3 method for class 'predmicror_omnibus_fit'  
residuals(object, ...)
```

```
## S3 method for class 'predmicror_omnibus_fit'  
predict(object, newdata = NULL, level = 0, ...)
```

```
## S3 method for class 'predmicror_omnibus_fit'  
logLik(object, ...)
```

```
## S3 method for class 'predmicror_omnibus_fit'  
AIC(object, ..., k = 2)
```

```
## S3 method for class 'predmicror_omnibus_fit'  
BIC(object, ...)
```

Arguments

x, object	A predmicror_omnibus_fit object.
...	Additional arguments passed to the underlying nlme method.
newdata	Optional data frame for prediction.
level	Prediction level passed to <code>stats::predict()</code> .
k	Penalty used by AIC().

Value

The value returned by the corresponding method.

RichardsNLM	<i>Richards no lag growth model</i>
-------------	-------------------------------------

Description

RichardsNLM function to fit the Richards no lag growth model to an incomplete microbial growth curve. Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

RichardsNLM(*t*, *Y0*, *Ymax*, *MUmax*, *m* = 1)

Arguments

<i>t</i>	is a numeric vector indicating the time of the experiment
<i>Y0</i>	is the natural logarithm of the initial microbial concentration ($\ln(N0)$) at time=0
<i>Ymax</i>	is the natural logarithm of the maximum concentration ($\ln(Nmax)$) reached by the microorganism
<i>MUmax</i>	is the maximum specific growth rate given in time units
<i>m</i>	is the shape parameter of the Richards model (default = 1)

Details

Model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the microbial concentration ($\ln(N(t))$) measured at time *t*.

Users should make sure that the microbial concentration input is entered in natural logarithm, $Y(t) = \ln(N(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez, <vcadavez@ipb.pt> and Ursula Gonzales-Barron, <ubarron@ipb.pt>

References

Richards JF (1959). "A flexible growth function for empirical use." *J Exp Bot*, **1**(10), 290-310.

Examples

```
## Example: Richards no lag model
library(gslNls)
data(growthnolag) # simulated data set.
initial_values <- list(Y0 = 0, Ymax = 22, MUmax = 1.7)
## Fitting the function to the experimental data
fit <- gsl_nls(lnN ~ RichardsNLM(Time, Y0, Ymax, MUmax),
  data = growthnolag,
  start = initial_values
)
summary(fit)
```

RossoFM

Rosso full growth model

Description

RossoFM function to fit the Rosso full growth model to complete microbial growth curve. Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
RossoFM(t, Y0, Ymax, MUmax, lag)
```

Arguments

t	is a numeric vector indicating the time of the experiment
Y0	is the natural logarithm of the initial microbial concentration ($\ln(N_0)$) at time=0
Ymax	is the natural logarithm of the maximum concentration ($\ln(N_{max})$) reached by the microorganism
MUmax	is the maximum specific growth rate given in time units
lag	is the duration of the lag phase in time units

Details

Model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the microbial concentration ($\ln(N(t))$) measured at time t.

Users should make sure that the microbial concentration input is entered in natural logarithm, $Y(t) = \ln(X(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez (<vcadavez@ipb.pt>) and Ursula Gonzales-Barron (<ubarron@ipb.pt>)

References

Rosso L, Bajard S, Flandrois JP, Lahellec C, Fournaud J, Veit P (1996). "Differential growth of *Listeria monocytogenes* at 4 and 8 °C: Consequences for the Shelf Life of Chilled Products." *Journal of Food Protection*, **59**(9), 944-949. ISSN 0362-028X, doi:10.4315/0362028X59.9.944.

Examples

```
## Example: Rosso full model
library(gsl_nls)
data(growthfull) # simulated data set.
initial_values <- list(Y0 = 0.04, Ymax = 21, MUmmax = 1.9, lag = 5.0) # define the initial values
fit <- gsl_nls(lnN ~ RossoFM(Time, Y0, Ymax, MUmmax, lag),
  data = growthfull,
  start = initial_values
)
summary(fit)
```

salmonella

Potential growth of Salmonella typhimurium on cooked chicken

Description

A dataset containing the specific growth rate of *Salmonella typhimurium* on cooked chicken

Usage

```
data(salmonella)
```

Format

A data frame with 21 rows and 3 variables:

Temp Temperature (°C)

GR Specific growth rate (log10/h)

sqrtGR Square root of the specific growth rate (GR)

References

Oscar TP (2002). "Development and validation of a tertiary simulation model for predicting the potential growth of *Salmonella typhimurium* on cooked chicken." *International Journal of Food Microbiology*, **76**(3), 177-190. ISSN 0168-1605, doi:10.1016/S01681605(02)000259.

```
validate_omnibus_leave_one_out
```

Validate an omnibus fit by leaving out one group

Description

Validate an omnibus fit by leaving out one group

Usage

```
validate_omnibus_leave_one_out(object, group_value, level = 0, ...)
```

Arguments

object	A <code>predmicror_omnibus_fit</code> object.
group_value	Group value to leave out.
level	Prediction level. Defaults to 0 for population-level prediction of the left-out group.
...	Additional arguments passed to <code>fit_omnibus()</code> during refitting.

Value

A list with the refitted model, validation data, predictions, residuals, bias factor, and accuracy factor.

```
WeibullM
```

Weibull inactivation model Mafart

Description

WeibullM inactivation model for microbial inactivation curve. Returns the model parameters estimated according to data collected in microbial inactivation experiments.

Usage

```
WeibullM(x, Y0, sigma, alpha)
```

Arguments

x	is a numeric vector indicating the heating time under a constant temperature of the experiment
Y0	is the natural logarithm of the initial (time=0) bacterial concentration (N0)
sigma	is the time of first decimal reduction
alpha	which is a shape parameter

Details

The model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the bacterial concentration ($\ln(N(t))$) measured at time t.

Users should make sure that the bacterial concentration input is entered in natural logarithm, $Y(t) = \ln(N(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Mafart P, Couvert O, Gaillard S, Leguerinel (2002). "On calculating sterility in thermal preservation methods: application of the Weibull frequency distribution model." *International Journal of Food Microbiology*, **72**, 107-113.

Examples

```
library(gslnls)
data(bixina)
initial_values <- list(Y0 = 5.75, sigma = 12.8, alpha = 2.4)
fit <- gsl_nls(lnN ~ WeibullM(Time, Y0, sigma, alpha),
  data = bixina,
  start = initial_values
)
summary(fit)

plot(lnN ~ Time, data = bixina)
lines(bixina$Time, predict(fit), col = "blue")
```

WeibullMM

Weibull inactivation modified model Mafart

Description

WeibullMM inactivation model for microbial inactivation curve. Returns the model parameters estimated according to data collected in microbial inactivation experiments.

Usage

```
WeibullMM(x, Y0, Yres, sigma, alpha)
```

Arguments

x	is a numeric vector indicating the heating time under a constant temperature of the experiment
Y0	is the log10 of the initial bacterial concentration (at time t=0)
Yres	is the log10 of the residual bacterial concentration (at the end of the experiment)
sigma	represents the time of the first decimal reduction concentration for the part of the population not belonging to Yres
alpha	is the shape parameter and allows to catch the curve concavity or convexity

Details

The model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the bacterial concentration (Y(t)) measured at time t.

Users should make sure to use the base 10 logarithm bacterial concentration (Y(t)) as input.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Mafart et al. (2005).

Examples

```
library(gslnls)
data(bixina)
initial_values <- list(Y0 = 5.6, Yres = 1, sigma = 2, alpha = 1)
bixina$logN <- log10(exp(bixina$lnN))
fit <- gsl_nls(logN ~ WeibullMM(Time, Y0, Yres, sigma, alpha),
  data = bixina,
  start = initial_values
)
summary(fit)

plot(logN ~ Time, data = bixina)
lines(bixina$Time, predict(fit), col = "blue")
```

WeibullPH

Weibull inactivation model Peleg and Huang

Description

WeibullPH inactivation model for microbial inactivation curve. Returns the model parameters estimated according to data collected in microbial inactivation experiments.

Usage

WeibullPH(t, Y0, k, alpha)

Arguments

t	is a numeric vector indicating the time of the experiment
Y0	is the natural logarithm of the initial (time=0) bacterial concentration
k	is the inactivation rate (ln units/time)
alpha	is the shape parameter

Details

The model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the bacterial concentration X(t) measured at time t.

Users should make sure that the bacterial concentration input is entered in natural logarithm, $Y(t) = \ln(X(t))$.

The following parameters can be estimated using Weibull function:

t: is heating time under a constant temperature

Y0: is the initial (time=0) bacterial counts in natural logarithm of the initial bacterial counts;

k: is the inactivation rate (ln units/time)

alpha: is the shape parameter of the survival curve

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Huang L (2009). "Thermal inactivation of *Listeria monocytogenes* in ground beef under isothermal and dynamic temperature conditions." *Journal of Food Engineering*, **90**(3), 380-387. ISSN 0260-8774, doi:[10.1016/j.jfoodeng.2008.07.011](https://doi.org/10.1016/j.jfoodeng.2008.07.011).

Examples

```

library(gslNls)
data(bixina)
initial_values <- list(Y0 = 6.0, k = 1.0, alpha = 0.2)
fit <- gsl_nls(lnN ~ WeibullPH(Time, Y0, k, alpha),
  data = bixina,
  start = initial_values
)
summary(fit)

plot(lnN ~ Time, data = bixina)
lines(bixina$Time, predict(fit), col = "blue")

```

ZwieteringFM

Zwietering full growth model

Description

ZwieteringFM function to fit the Zwietering full growth model to a complete microbial growth curve. Returns the model parameters estimated according to data collected in microbial growth experiments.

Usage

```
ZwieteringFM(t, Y0, Ymax, MUmex, lag)
```

Arguments

t	is a numeric vector indicating the time of the experiment
Y0	is the natural logarithm of the initial microbial concentration ($\ln(N_0)$) at time=0
Ymax	is the natural logarithm of the maximum concentration ($\ln(N_{max})$) reached by the microorganism
MUmex	is the maximum specific growth rate given in time units
lag	is the duration of the lag phase in time units

Details

Model's inputs are:

t: time, assuming time zero as the beginning of the experiment.

Y(t): the natural logarithm of the microbial concentration ($\ln(N(t))$) measured at time t.

Users should make sure that the microbial concentration input is entered in natural logarithm, $Y(t) = \ln(X(t))$.

Value

A numeric vector with the fitted values

Author(s)

Vasco Cadavez <vcadavez@ipb.pt> and Ursula Gonzales-Barron <ubarron@ipb.pt>

References

Zwietering MH, Jongenburger I, Rombouts FM, van't Riet K (1990). "Modeling of the Bacterial Growth Curve." *Applied and Environmental Microbiology*, **56**(6), 1875-1881. ISSN 0099-2240.

Examples

```
## Example: Zwietering full model
library(gsl_nls)
data(growthfull) # simulated data set.
initial_values <- list(Y0 = 0, Ymax = 22, MUmax = 1.7, lag = 5) # define the initial values
fit <- gsl_nls(lnN ~ ZwieteringFM(Time, Y0, Ymax, MUmax, lag),
  data = growthfull,
  start = initial_values
)
summary(fit)
```

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